

CURRICULUM VITAE

Roy Parker

Howard Hughes Medical Institute
Department of Biochemistry
University of Colorado Boulder
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EDUCATION:

- Postdoctoral: University of Massachusetts Medical School, 1/88-9/89
(Laboratory of Dr. Allan Jacobson)
University of California at San Diego, 10/86-12/87
(Laboratory of Dr. Michael Yaffe)
University of California at San Francisco, 4/85-9/86
(Laboratory of Dr. Christine Guthrie)
- Ph.D.: University of California at San Francisco, 1985 (Genetics)
- B.S.: Carnegie Mellon University, Pittsburgh, PA, 1979 (Chemistry)

ACADEMIC POSITIONS:

- 1989-1995 Assistant Professor
Department of Molecular and Cellular Biology, University of Arizona
- 1994-1997 Assistant Investigator
Howard Hughes Medical Institute
- 1995-1998 Associate Professor
Department of Molecular and Cellular Biology, University of Arizona
- 1997-2002 Associate Investigator
Howard Hughes Medical Institute
- 1998-2001 Professor
Department of Molecular and Cellular Biology, University of Arizona
- 2001-2012 Regents' Professor
Department of Molecular and Cellular Biology, University of Arizona
- 2002-present Investigator
Howard Hughes Medical Institute
- 2012-present Cech-Leinwand Endowed Chair of Biochemistry, and Professor, Department of Biochemistry, University of Colorado Boulder
- 2018-present Distinguished Professor, Department of Biochemistry, University of Colorado Boulder
- 2020-present Executive Director, BioFrontiers Institute, University of Colorado Boulder

OTHER PROFESSIONAL ACTIVITIES:

1996-1997	Member, Defense Science Study Group
1996-2005	Consultant, Institute for Defense Analyses
1997	Co-Chair, Gordon Conference on Nucleic Acids
1999	Co-Organizer, CSHL Conference on RNA Processing
1997-2000	Member, NIH Molecular Biology Study Section
1999-2000	Chair, NIH Molecular Biology Study Section
2004	Co-Organizer FASEB Conference on Post-Transcriptional Control
2014-2021	Friedrich Miescher Institute for Biomedical Research, Scientific Advisory Board
2019-2022	Consultant for Third Rock Ventures
2019-2022	Co-Founder of Faze Medicines
2023- present	Scientific Advisory Board, Center for RNA Biomedicine, University of Michigan, Ann Arbor

HONORS:

Regents' Fellowship (1979)
Searle Scholarship (1990)
Regents' Professor (2003)
Galileo Fellow (2003)
NIH Merit Award (2004)
President, RNA Society
Member, American Academy of Arts & Sciences (2010)
Member, National Academy of Sciences (2012)
Executive Director, BioFrontiers Institute, University of Colorado Boulder, (2020)
Robert Sterns Award for Extraordinary Service (2021)
Over 30 Keynote and Distinguished Lectures (2001-present)

EDITOR & EDITORIAL BOARDS:

Molecular and Cellular Biology 1994 - 1997
Science Board of Reviewing Editors 1997 – 1999, 2001 - 2004
RNA 1999 – present
Nucleic Acids Research 2004 – 2017
Editor, Journal of Cell Biology 2008 – 2017
Cell 2009 – present
Current Biology 2014 – present

PUBLICATIONS:**ORIGINAL RESEARCH PAPERS**

1. Jones, E.W., Zubenko, G.S. and **R.R. Parker**. (1982) PEP4 gene function is required for expression of several vacuolar hydrolases in *Saccharomyces cerevisiae*. **Genetics** 102:665-677. [PMCID: PMC1201965](#)
2. **Parker, R.** and C. Guthrie. (1985) A point mutation in the conserved hexanucleotide at a yeast 5' splice site uncouples recognition, cleavage and ligation. **Cell** 41:107-118.
3. Cellini, A., **Parker, R.**, McMahon, J., Guthrie, C. and J. Rossi. (1986) Activation of a cryptic TACTAAC box in the *Saccharomyces cerevisiae*. **Molecular and Cellular Biology** 6:1571-1578. [PMCID: PMC367683](#)

Original Research Papers (continued)

4. Vijayraghavan, U., **Parker, R.**, Tamm, J., Iimura, Y., Rossi, J., Abelson, J.A. and C. Guthrie. (1986) Mutations in conserved intron sequences affect multiple steps in the yeast splicing pathway, particularly assembly of the spliceosome. **EMBO Journal** 5:1683-1695. [PMCID: PMC1166995](#)
5. Couto, J.R., Tamm, J., **Parker, R.** and C. Guthrie. (1987) A trans-acting suppressor restores splicing of a yeast intron with a branch point mutation. **Genes & Development** 1:445-455.
6. **Parker, R.**, Siliciano, P.G. and C. Guthrie. (1987) Recognition of the TACTAAC box during mRNA splicing in yeast involves base-pairing to the U2-like snRNA. **Cell** 49:229-239.
7. Ruohola, H., Baker, S.M., **Parker, R.** and T. Platt. (1988) Orientation-dependent function of a short CYC1 DNA fragment in directing mRNA 3' end formation in yeast. **Proceeding of the National Academy of Science** 85:5041-5045. [PMCID: PMC281683](#)
8. **Parker, R.**, Simmons, T., Shuster, E., Siliciano, P.G. and C. Guthrie. (1988) Genetic analysis of small nuclear RNAs in *Saccharomyces cerevisiae*: viable sextuple mutant. **Molecular and Cellular Biology** 8:3150-3159. [PMCID: PMC363543](#)
9. **Parker, R.** and A. Jacobson. (1990) Translation and a 42-nucleotide segment within the coding region of the mRNA encoded by the *MATα1* gene are involved in promoting rapid mRNA decay in yeast. **Proceeding of the National Academy of Science** 87:2780-2784. [PMCID: PMC53774](#)
10. Herrick, D., **Parker, R.** and A. Jacobson. (1990) Identification and comparison of stable and unstable mRNAs in *Saccharomyces cerevisiae*. **Molecular and Cellular Biology** 10:2269-2284. [PMCID: PMC360574](#)
11. Muhlrad, D., Hunter, R. and **R. Parker**. (1992) A rapid method for the localized mutagenesis of yeast genes. **Yeast** 8:79-82.
12. Heaton, B., Decker, C., Muhlrad, D., Donahue, J., Jacobson, A. and **R. Parker**. (1992) Analysis of chimeric mRNAs derived from STE3 mRNA identifies multiple regions within yeast mRNAs that regulate mRNA decay. **Nucleic Acids Research** 20:5365-5373. [PMCID: PMC334343](#)
13. Muhlrad, D. and **R. Parker**. (1992) Mutations affecting stability and deadenylation of the yeast MFA2 transcript. **Genes & Development** 6:2100-2111.
14. **Parker, R.** and P.G. Siliciano. (1993) Evidence for a non-Watson Crick interaction between the first and last nucleotides of a nuclear pre-mRNA intron. **Nature** 361:660-662.
15. Decker, C.J. and **R. Parker**. (1993) A turnover pathway for both stable and unstable mRNAs in yeast: evidence for a requirement for deadenylation. **Genes & Development** 7:1632-1643.

Original Research Papers (continued)

16. Caponigro, G., Muhlrad, D. and **R. Parker**. (1993) A small segment of the MAT α 1 transcript promotes mRNA decay in yeast: a stimulatory role for rare codons. **Molecular and Cellular Biology** 13:5141-5148. PMCID: PMC360202
17. Beelman, C.A. and **R. Parker**. (1994) Differential effects of translational inhibition in cis and in trans on the decay of the unstable yeast MFA2 mRNA. **Journal of Biological Chemistry** 269:9687-9692.
18. Muhlrad, D., Decker, C.J. and **R. Parker**. (1994) Deadenylation of the unstable mRNA encoded by the yeast MFA2 gene leads to decapping followed by 5' \rightarrow 3' digestion of the transcript. **Genes & Development** 8:855-866.
19. Muhlrad, D. and **R. Parker**. (1994) Premature translational termination triggers mRNA decapping. **Nature** 370:578-581.
20. Muhlrad, D., Decker, C.J. and **R. Parker**. (1995) Turnover mechanisms of the stable PGK1 mRNA in yeast. **Molecular and Cellular Biology** 15:2145-2156. PMCID: PMC230442
21. Caponigro, G. and **R. Parker**. (1995) Multiple functions of the Poly(A) binding protein in mRNA decapping and deadenylation in yeast. **Genes & Development** 9:2421-2432.
22. Mandart, E. and **R. Parker**. (1995) Effects of mutations in the RNA14, RNA15 and PAP1 genes on polyadenylation *in vivo*. **Molecular and Cellular Biology** 15:6979-6986. PMCID: PMC230953
23. Hatfield, L., Beelman, C. A., Stevens A. and **R. Parker**. (1996) Mutations in trans-acting factors affecting mRNA decapping in *Saccharomyces cerevisiae*. **Molecular and Cellular Biology** 16:5830-5838. PMCID: PMC231584
24. Beelman, C. A., Stevens A., Caponigro G., LaGrandeur, T.E., Hatfield L., Fortner, D. and **R. Parker**. (1996) An essential component of the decapping enzyme required for normal rates of mRNA decay in yeast. **Nature** 382:642-646.
25. Caponigro, G. and **R. Parker**. (1996) mRNA turnover in yeast promoted by the MAT α 1 instability element. **Nucleic Acids Research** 24:4304-4312. PMCID: PMC146253
26. Olivas, W. M., Muhlrad, D. and **R. Parker**. (1997) Analysis of the yeast genome: identification of new non-coding and small ORF-containing RNAs. **Nucleic Acids Research** 25:4619-4625. PMCID: PMC147069
27. Anderson, J.S.J. and **R. Parker**. (1998) The 3' to 5' degradation of yeast mRNAs is a general mechanism for mRNA turnover that requires the SKI2 DE VH box protein and 3' to 5' exonucleases of the exosome complex. **EMBO Journal** 17:1497-1506. PMCID: PMC1170497
28. LaGrandeur, T.E. and **R. Parker**. (1998) Isolation and characterization of Dcp1p, the yeast mRNA decapping enzyme. **EMBO Journal** 17:1487-1496. PMCID: PMC1170496

Original Research Papers (continued)

29. LaGrandeur, T. and **R. Parker**. (1999) The *cis* acting sequences responsible for the differential decay of the unstable MFA2 and stable PGK1 transcripts in yeast includes the context of the translational start codon. **RNA** 5:420-433. [PMCID: PMC1369770](#)
30. Tharun, S. and **R. Parker**. (1999) Analysis of mutations in the yeast mRNA decapping enzyme. **Genetics** 151:1273-1285. [PMCID: PMC1460575](#)
31. Schwartz, D.C. and **R. Parker**. (1999) Mutations in translation initiation factors lead to increased rates of deadenylation and decapping of mRNAs in *Saccharomyces cerevisiae*. **Molecular and Cellular Biology** 19:5247-5256. [PMCID: PMC84368](#)
32. Dunckley, T. and **R. Parker**. (1999) The DCP2 protein is required for mRNA decapping in *Saccharomyces cerevisiae* and contains a functional MutT motif. **EMBO Journal** 18: 5411-5422. [PMCID: PMC1171610](#)
33. Muhlrad, D. and **R. Parker**. (1999) Recognition of yeast mRNAs as "nonsense containing" leads to both inhibition of mRNA translation and mRNA degradation: Implications for the control of mRNA decapping. **Molecular Biology of the Cell** 10: 3971-3978. [PMCID: PMC25692](#)
34. Muhlrad, D. and **R. Parker**. (1999) Aberrant mRNAs with extended 3' UTRs are substrates for rapid degradation by mRNA surveillance. **RNA** 5:1299-1307. [PMCID: PMC1369852](#)
35. van Hoof, A., Lennertz, P. and **R. Parker**. (2000) Yeast exosome mutants accumulate 3'-extended polyadenylated forms of U4 small nuclear RNA and small nucleolar RNAs. **Molecular and Cellular Biology** 20:441-452. [PMCID: PMC85098](#)
36. van Hoof, A., Lennertz, P. and **R. Parker**. (2000) Three conserved members of the RNase D family have unique and overlapping functions in the processing of 5S, 5.8S, U4, U5, RNase MRP and RNase P RNAs in yeast. **EMBO Journal** 19: 1357-1365. [PMCID: PMC305676](#)
37. Tharun, S., He, W., Mayes, A.E., Lennertz, P., Beggs, J.D. and **R. Parker**. (2000) Yeast Sm-like proteins function in mRNA decapping and decay. **Nature** 404:515-518.
38. Jacobs Anderson, J.S. and **R. Parker**. (2000) Computational identification of *cis*-acting elements affecting post-transcriptional control of gene expression in *Saccharomyces cerevisiae*. **Nucleic Acids Research** 28:1604-1617. [PMCID: PMC102784](#)
39. Schwartz, D.C. and **R. Parker**. (2000) mRNA decapping in yeast requires dissociation of the cap binding protein, eukaryotic translation initiation factor 4E. **Molecular and Cellular Biology** 20:7933-7942. [PMCID: PMC86404](#)
40. van Hoof, A., Staples, R.R., Baker, R.E. and **R. Parker**. (2000) Function of the Ski4p (Csl4p) and Ski7p proteins in 3'-to-5' degradation of mRNA. **Molecular and Cellular Biology** 20:8230-8243. [PMCID: PMC86432](#)

Original Research Papers (continued)

41. Olivas, W. and **R. Parker**. (2000) The Puf3 protein is a transcript-specific regulator of mRNA degradation in yeast. **EMBO Journal** 19:6602-6611. [PMCID: PMC305854](#)
42. Dunckley T., Tucker, M. and **R. Parker**. (2001) Two related proteins, Edc1p and Edc2p, stimulate mRNA decapping in *Saccharomyces cerevisiae*. **Genetics** 157:27-37. [PMCID: PMC1461477](#)
43. Tucker, M., Valencia-Sanchez M.A., Staples R.R., Chen J., Denis C.L. and **R. Parker**. (2001) The transcription factor associated proteins Ccr4 and Caf1 proteins are components of the major cytoplasmic mRNA deadenylase in *Saccharomyces cerevisiae*. **Cell** 104:377-386.
44. Hilleren, P. and **R. Parker**. (2001) Defects in the mRNA export factors Rat7p, Gle1p, Mex67p and Rat8p cause hyperadenylation during 3' end formation of nascent transcripts. **RNA** 7:753-764. [PMCID: PMC1370127](#)
45. He, W. and **R. Parker**. (2001) The yeast cytoplasmic Lsm1/Pat1p complex protects mRNA 3' termini from partial degradation. **Genetics** 158:1445-1455. [PMCID: PMC1461746](#)
46. Cao, D. and **R. Parker**. (2001) Computational modeling of eukaryotic mRNA turnover. **RNA** 7:1192-1212. [PMCID: PMC1370166](#)
47. Tharun, S. and **R. Parker**. (2001) Targeting an mRNA for decapping: displacement of translation factors and association of the Lsm1p-7p complex on deadenylated yeast mRNAs. **Molecular Cell** 8:1075-1083.
48. Coller, J.M., Tucker, M., Sheth, U., Valencia-Sanchez, M.A. and **R. Parker**. (2001) The DEAD box helicase, Dhh1p, functions in mRNA decapping and interacts with both the decapping and deadenylase complexes. **RNA** 7:1717-1727. [PMCID: PMC1370212](#)
49. Hilleren, P., McCarthy, T., Rosbash, M., **Parker, R.** and T.H. Jensen. (2001) Quality control of mRNA 3'-end processing is linked to the nuclear exosome. **Nature** 413:538-542.
50. Tucker, M., Staples, R.R., Valencia-Sanchez, M.A., Muhlrad, D. and **R. Parker**. (2002) Ccr4p is the catalytic sub-unit of a Ccr4/Pop2p/Notp mRNA deadenylase complex in *Saccharomyces cerevisiae*. **EMBO Journal** 21:1427-1436. [PMCID: PMC125913](#)
51. van Hoof, A., Frischmeyer, P.A., Dietz, H.C. and **R. Parker**. (2002) Exosome-mediated recognition and degradation of mRNAs lacking a termination codon. **Science** 295:2262-2264.
52. Frischmeyer, P.A., van Hoof, A., O'Donnell, K., Guerriero, A.L., **Parker, R.** and H.C. Dietz. (2002) An mRNA surveillance mechanism that eliminates transcripts lacking termination codons. **Science** 295:2258-2261.
53. Steiger, M., Carr-Schmid, A., Schwartz, D.C., Kiledjian, M. and **R. Parker**. (2003) Analysis of recombinant yeast decapping enzyme. **RNA** 9:231-238. [PMCID: PMC1370389](#)

Original Research Papers (continued)

54. Schwartz, D., Decker, C.J. and **R. Parker**. (2003) The enhancer of decapping proteins, Edc1p and Edc2p, bind RNA and stimulate activity of the decapping enzyme. **RNA** 9:239-251. [PMCID: PMC1370390](#)
55. Sheth, U. and **R. Parker**. (2003) Decapping and decay of messenger RNA occur in cytoplasmic processing bodies. **Science** 300:805-808. [PMCID: PMC1876714](#)
56. Cao, D. and **R. Parker**. (2003) Computational modeling and experimental analysis of nonsense-mediated decay in yeast. **Cell** 113:533-545.
57. Hilleren, P.J. and **R. Parker**. (2003) Cytoplasmic degradation of splice-defective pre-mRNAs and intermediates. **Molecular Cell** 12:1453-1465.
58. She, M., Decker, C.J., Sundramurthy, K., Liu, Y., Chen, N., **Parker, R.** and H. Song. (2004) Crystal structure of Dcp1p and its functional implications in mRNA decapping. **Nature Structural & Molecular Biology** 11:249-256. [PMCID: PMC2040073](#)
59. Kshirsagar, M. and **R. Parker**. (2004) Identification of Edc3p as an enhancer of mRNA decapping in *Saccharomyces cerevisiae*. **Genetics** 166:729-739. [PMCID: PMC1470743](#)
60. Baker, K.E., Coller, J.M. and **R. Parker**. (2004) The yeast Apq12 protein affects nucleocytoplasmic mRNA transport. **RNA** 10:1352-1358. [PMCID: PMC1370622](#)
61. Cheng, Z., Liu, Y., Wang, C., **Parker R.** and H. Song. (2004) Crystal structure of Ski8p, a WD-repeat protein with dual roles in mRNA metabolism and meiotic recombination. **Protein Science** 13:2673-2684. [PMCID: PMC2001155](#)
62. Chen, N., Walsh, M.A., Liu, Y., **Parker, R.** and H. Song. (2005) Crystal structures of human DcpS in ligand-free and m⁷GDP-bound forms suggest a dynamic mechanism for scavenger mRNA decapping. **Journal of Molecular Biology** 347:707-718.
63. Muhlrad, D. and **R. Parker**. (2005) The yeast EDC1 mRNA undergoes deadenylation-independent decapping stimulated by Not2p, Not4p, and Not5p. **EMBO Journal** 24:1033-1045. [PMCID: PMC554118](#)
64. Teixeria, D., Sheth, U., Valencia-Sanchez, MA, Brengues, M. and **R. Parker**. (2005) Processing bodies require RNA for assembly and contain non-translating mRNAs. **RNA** 11:371-382. [PMCID: PMC1370727](#)
65. Tharun, S., Muhlrad, D., Chowdhury, A. and **R. Parker**. (2005) Mutations in the *Saccharomyces cerevisiae* LSM1 gene that affect mRNA decapping and 3' end protection. **Genetics** 170:33-46. [PMCID: PMC1449704](#)
66. Cheng, Z., Coller, J., **Parker, R.** and H. Song. (2005) Crystal structure and functional analysis of DEAD-box protein Dhh1p. **RNA** 11:1258-1270. [PMCID: PMC1370809](#)
67. Wilson, M.A., Meaux, S., **Parker, R.** and A. van Hoof. (2005) Genetic interactions between [PSI⁺] and nonstop mRNA decay affect phenotypic variation **Proceeding of the National Academy of Science** 102:10244-10249. [PMCID: PMC1173365](#)

Original Research Papers (continued)

68. Coller, J. and **R. Parker**. (2005) General translational repression by activators of mRNA decapping. **Cell** 122:875-886. [PMCID: PMC1853273](#)
69. Brengues, M., Teixeria, D. and **R. Parker**. (2005) Movement of eukaryotic mRNAs between polysomes and cytoplasmic processing bodies. **Science** 310:486-489. [PMCID: PMC1863069](#)
70. Liu, J., Valencia-Sanchez, M.A., Hannon, G.J. and **Parker, R.** (2005). MicroRNA-dependent localization of targeted mRNAs to mammalian P-bodies. **Nature Cell Biology** 7:719-723. [PMCID: PMC1855297](#)
71. Liu, J., Rivas, F.V., Wohlschlegel, J., Yates III, J.R., **Parker, R.** and G.J. Hannon. (2005) A role for the P-body component GW182 in microRNA function. **Nature Cell Biology** 7:1261-1266. [PMCID: PMC1804202](#)
72. Beliakova-Bethell, N., Beckham, C., Giddings Jr., T.H., Winey, M., **Parker, R.** and S. Sandmeyer. (2006) Virus-like particles of the Ty3 retrotransposon assemble in association with P-body components. **RNA** 12:94-101. [PMCID: PMC1370889](#)
73. She, M., Decker, C.J., Chen, N., Tumati, S., **Parker, R.** and H. Song. (2006) Crystal structure and functional analysis of Dcp2p from *Schizosaccharomyces pombe*. **Nature Structural and Molecular Biology** 13:63-70. [PMCID: PMC1952686](#)
74. Doma, M.K. and **R. Parker**. (2006) Endonucleolytic cleavage of eukaryotic mRNAs with stalls in translation elongation. **Nature** 440:561-564. [PMCID: PMC1839849](#)
75. Sheth, U. and **R. Parker**. (2006) Targeting of aberrant mRNAs to cytoplasmic processing bodies. **Cell** 125:1095-1109. [PMCID: PMC1858659](#)
76. Hilgers, V., Teixeira, D. and **R. Parker**. (2006). Translation-independent inhibition of mRNA deadenylation during stress in *Saccharomyces cerevisiae*. **RNA** 12:1835-1845. [PMCID: PMC1581975](#)
77. Segal, S.P., Dunckley, T.C. and **R. Parker**. (2006) Sbp1p affects translational repression and decapping in *Saccharomyces cerevisiae*. **Molecular and Cellular Biology** 26:5120-5130. [PMCID: PMC1489156](#)
78. Baker, K.E. and **R. Parker**. (2006) Conventional 3' end formation is not required for NMD substrate recognition in *Saccharomyces cerevisiae*. **RNA** 12:1441-1445.
79. Barbee, S.A., Estes, P.S., Cziko, A-M., Hillebrand, J., Luedeman, R.A., Coller, J.M., Johnson, N., Howlett, I.C., Geng, C., Ueda, R., Brand, A.H., Newbury, S.F., Wilhelm, J.E., Levine, R.B., Nakamura, A., **Parker, R.**, and M. Ramaswami. (2006) Staufen- and FMRP-containing neuronal RNPs are structurally and functionally related to somatic P bodies. **Neuron** 52:997-1009. [PMCID: PMC1955741](#)

Original Research Papers (continued)

80. Thompson, D. M. and **R. Parker**. (2007) Cytoplasmic decay of intergenic transcripts in *Saccharomyces cerevisiae*. **Molecular and Cellular Biology** 27:92-101. [PMCID: PMC1800667](#)
81. Cheng, Z., Muhlrad, D., Lim, M.K., **Parker, R.** and H. Song (2007) Structural and functional insights into the human Upf1 helicase core. **EMBO Journal** 26:253-64. [PMCID: PMC1782376](#)
82. Teixeira, D. and **R. Parker**. (2007) Analysis of P-body assembly in *Saccharomyces cerevisiae*. **Molecular Biology of the Cell** 18:2274-2287. [PMCID: PMC1877105](#)
83. Brengues, M. and **R. Parker**. (2007) Accumulation of polyadenylated mRNA, Pab1p, eIF4E, and eIF4G with P-bodies in *Saccharomyces cerevisiae*. **Molecular Biology of the Cell** 18:2592-2602. [PMCID: PMC1924816](#)
84. Beckham, C.J., Light, H.R., Nissan, T.A., Ahlquist, P., **Parker, R.**, and A. Nouiry. (2007) Interactions between brome mosaic virus RNAs and cytoplasmic processing bodies. **Journal of Virology** 81:9759-9768. [PMCID: PMC2045432](#)
85. Decker, C.J., Teixeira, D., and **R. Parker**. (2007) Edc3p and a glutamine/asparagine-rich domain of Lsm4p function in processing body assembly in *Saccharomyces cerevisiae*. **Journal of Cell Biology** 179:437-449. [PMCID: PMC2064791](#)
86. Pilkington, G.R., and **R. Parker**. (2008) Pat1 contains distinct functional domains that promote P-body assembly and activation of decapping. **Molecular and Cellular Biology** 28:1298-1312. [PMCID: PMC2258743](#)
87. Beckham, C.J., Hilliker, A., Cziko, A.-M., Noueiry, A., Ramaswami, M., and **R. Parker**. (2008) The DEAD-Box RNA helicase Ded1p affects and accumulates in *Saccharomyces cerevisiae* P-bodies. **Molecular Biology of the Cell** 19:984-993. [PMCID: PMC2262982](#)
88. She, M., Decker, C.J., Svergun, D.I., Round, A., Chen, N., Muhlrad, D., **Parker, R.**, and H. Song. (2008) Structural basis of Dcp2 recognition and activation by Dcp1. **Molecular Cell** 29:337-349. [PMCID: PMC2323275](#)
89. Nissan, T. and **R. Parker**. (2008) Computational analysis of miRNA-mediated repression of translation: Implications for models of translation initiation inhibition. **RNA** 14:1480-1491. [PMCID: PMC2491470](#)
90. Ling, S.H., Decker, C.J., Walsh, M.A., She, M., **Parker, R.** and H. Song (2008) Crystal structure of human Edc3 and its functional implications. **Molecular and Cellular Biology** 28:5965-5976. [PMCID: PMC2547010](#)
91. Pedro-Segura, E., Vergara, S.V., Rodríguez-Navarro, S., **Parker, R.**, Thiele, D.J. and S. Puig (2008) The Cth2 ARE-binding protein recruits the Dhh1 helicase to promote the decay of succinate dehydrogenase SDH4 mRNA in response to iron deficiency. **Journal of Biological Chemistry** 283:28527-28535. [PMCID: PMC2568921](#)

Original Research Papers (continued)

92. Thompson, D.M., Lu, C., Green, P.J. and **R. Parker**. (2008) tRNA cleavage is a conserved response to oxidative stress in eukaryotes. *RNA* 14:2095-2103. [PMCID: PMC2553748](#)
93. Buchan, J.R., Muhlrad, D. and **R. Parker**. (2008) P-bodies promote stress granule assembly in *Saccharomyces cerevisiae*. *Journal of Cell Biology* 183:441-455. [PMCID: PMC2575786](#)
94. Balagopal, V. and **R. Parker**. (2009) Sml1 modulates mRNA decay and Dhh1 function in *Saccharomyces cerevisiae*. *Genetics* 181:93-103. [PMCID: PMC2621192](#)
95. Thompson, D. and **R. Parker**. (2009) The RNase Rny1p cleaves tRNAs and promotes cell death during oxidative stress in *Saccharomyces cerevisiae*. *Journal of Cell Biology* 185:43-50. [PMCID: PMC2700514](#)
96. Chekulaeva, M., Filipowicz, W. and **R. Parker**. (2009) Multiple independent domains of dGW182 function in miRNA-mediated repression in *Drosophila*. *RNA* 15:794-803. [PMCID: PMC2673071](#)
97. Passos, D.O., Doma, M.K., Shoemaker, C.J., Muhlrad, D., Green, R., Weissman, J., Hollien, J. and **R. Parker**. (2009) Analysis of Dom34 and its function in no-go decay. *Molecular Biology of the Cell* 20:3025-3032. [PMCID: PMC2704154](#)
98. Luhtala, N. and **R. Parker**. (2009) LSM1 over-expression in *Saccharomyces cerevisiae* depletes U6 snRNA levels. *Nucleic Acids Research* 37:5529-5536. [PMCID: PMC2760792](#)
99. Cziko, A.-M.J., McCann, C.T., Howlett, I.C., Barbee, S.A., Duncan, R.P., Luedemann, R., Zarnescu, D., Zinsmaier, K.E., **Parker, R.R.** and M. Ramaswami. (2009) Genetic modifiers of *dfmr1* encode RNA granule components in *Drosophila*. *Genetics* 182:1051-1060. [PMCID: PMC2728847](#)
100. Harigaya, Y., Jones, B.N., Muhlrad, D., Gross, J.D. and **R. Parker**. (2010) Identification and analysis of the interaction between Edc3 and Dcp2 in *Saccharomyces cerevisiae*. *Molecular and Cellular Biology* 30:1446-1456. [PMCID: PMC2832485](#)
101. Yoon, J-H., Choi, E-J. and **R. Parker**. (2010) Dcp2 phosphorylation by Ste20 modulates stress granule assembly and mRNA decay in *Saccharomyces cerevisiae*. *Journal of Cell Biology* 189:813-827. [PMCID: PMC2878948](#)
102. Swisher K.D. and **R. Parker**. (2010) Localization to, and Effects of Pbp1, Pbp4, Lsm12, Dhh1, and Pab1 on Stress Granules in *Saccharomyces cerevisiae*. *PLOS ONE* 5:e10006. [PMCID: PMC2848848](#)
103. Nissan, T., Rajyaguru, P., She, M., Song, H. and **R. Parker**. (2010) Decapping activators in *Saccharomyces cerevisiae* act by multiple mechanisms. *Molecular Cell* 39:773-783. [PMCID: PMC2946179](#)
104. Chen, L., Muhlrad, D., Hauryliuk, V., Cheng, Z., Lim, M.K., Shyp, V., **Parker, R.** and H. Song. (2010) Structure of the Dom34-Hbs1 complex and implications for no-go decay. *Nature Structural & Molecular Biology* 17:1233-1240. [PMCID: PMC3682582](#)

Original Research Papers (continued)

105. Hillebrand, J., Pan, K., Kokaram, A., Barbee, S., **Parker, R.** and M. Ramaswami. (2010) The Me31B DEAD-box helicase localizes to postsynaptic foci and regulates expression of a CaMKII reporter mRNA in dendrites of *Drosophila* olfactory projection neurons. **Frontiers in Neural Circuits** 4:121 [PMCID: PMC3024558](#)
106. Chekulaeva, M., **Parker, R.** and W. Filipowicz. (2010) The GW/WG repeats of *Drosophila* GW182 function as effector motifs for miRNA-mediated repression. **Nucleic Acids Research** 38:6673-6683. [PMCID: PMC3965232](#)
107. Buchan, J.R., Yoon, J-H. and **R. Parker**. (2011) Stress-specific composition, assembly and kinetics of Stress granules in *Saccharomyces cerevisiae*. **Journal of Cell Science** 124:228-239. [PMCID: PMC3010191](#)
108. Balagopal, V. and **R. Parker**. (2011) Stm1 modulates translation after 80S formation in *Saccharomyces cerevisiae*. **RNA** 17:835-842. [PMCID: PMC3078733](#)
109. Hilliker, A., Gao, Z., Jankowsky, E. and **R. Parker**. (2011) The DEAD-box protein Ded1 modulates translation by the formation and resolution of an eIF4F-mRNA complex. **Molecular Cell** 43:962-972. [PMCID: PMC3268518](#)
110. McCann, C., Holohan E.E., Das, S., Dervan, A., Larkin, A., Lee, J.A., Rodrigues, V., **Parker, R.** and M. Ramaswami. (2011) The Ataxin-2 protein is required for microRNA function and synapse-specific long-term olfactory habituation. **Proceeding of the National Academy of Science** 108: E655-662. [PMCID: PMC3169144](#)
111. Chekulaeva, M., Mathys, H., Zipprich, J.T., Attig, J., Colic, M., **Parker, R.** and W. Filipowicz. (2011) miRNA repression involves GW182-mediated recruitment of CCR4-NOT through conserved W-containing motifs. **Nature Structural & Molecular Biology** 18:1218-1226. [PMCID: PMC385283](#)
112. Swisher, K. and **R. Parker**. (2011) Interactions between Upf1 and the decapping factors Edc3 and Pat1 in *Saccharomyces cerevisiae*. **PLOS ONE** 6: e26547 [PMCID: PMC3204985](#)
113. Rajyaguru, P., She, M. and **R. Parker**. (2012) Scd6 targets eIF4G to repress translation: RGG motif proteins as a class of eIF4G-binding proteins. **Molecular Cell** 45:244-254. [PMCID: PMC3277450](#)
114. Harigaya, Y. and **R. Parker**. (2012) Global analysis of mRNA decay intermediates in *Saccharomyces cerevisiae*. **Proceedings of the National Academy of Sciences** 109:11764-11769. [PMCID: PMC3406813](#)
115. **Parker, R.** (2012) Skill Development in Graduate Education. **Molecular Cell** 46:377-361. [PMCID: PMC3873173](#)
116. Luhtala, N. and **R. Parker**. (2012) Structure-function analysis of Rny1 in tRNA cleavage and growth inhibition. **PLOS ONE** 7:e41111. Epub 2012 Jul 19. [PMCID: PMC3400635](#)

Original Research Papers (continued)

117. Lai, T., Cho, H., Liu, Z., Bowler, M.W., Piao, S., **Parker, R.**, Kim, Y.K. and H. Song. (2012) Structural basis of PNRC2-mediated link between mRNA surveillance and decapping. **Structure** 20:2025-2037. [PMCID: PMC2323275](#)
118. Mitchell, S.F., Jain, S., She, M. and **R. Parker**. (2013) Global Analysis of Yeast mRNPs. **Nature Structural and Molecular Biology**, 20:127-133. [PMCID: PMC3537908](#)
119. Buchan, J.R., Kolaitis, R-M., Taylor, J.P and **R. Parker**. (2013) Eukaryotic stress granules are cleared by autophagy and Cdc48/VCP function. **Cell** 153:1461-1474. [PMCID: PMC3760148](#)
120. Sudhakaran, I.P., Hillebrand, J., Dervan, A., Das, S., Holohan, E.E., Hülsmeier, J., Sarov, M., **Parker, R.**, VijayRaghavan, K. and M. Ramaswami. (2014) FMRP and Ataxin-2 function together in long-term olfactory habituation and neuronal translational control. **Proceedings of the National Academy of Science** 111(1):E99-E108. [PMCID: PMC3890871](#)
121. Wu, D., Muhlrad, D., Bowler, M.W., Liu, Z., **Parker, R.** and H. Song. (2014) Lsm2 and Lsm3 bridge the interaction of the Lsm1-7 complex with Pat1 for decapping activation. **Cell Research** 24:233-246. [PMCID: PMC3915908](#)
122. Walters, R.W., Shumilin, I.A., Yoon, J-H., Minor, W. and **R. Parker**. (2014) Edc3 function in yeast and mammals is modulated by interaction with NAD-related compounds. **G3:Genes, Genomes, Genetics** 4:613-622. [PMCID: PMC4059234](#)
123. Decker, C.J. and **R. Parker**. (2014) Analysis of Double-Stranded RNA from Microbial Communities Identifies Double-Stranded RNA Virus-Like Elements. **Cell Reports** 7:898-906. [PMCID: PMC41117469](#)
124. Shukla, S. and **R. Parker**. (2014) Quality control of assembly-defective U1 snRNAs by decapping and 5'-to-3' exonucleolytic digestion. **Proceedings of the National Academy of Science** 111(32): E3277-E3286. [PMCID: PMC4136611](#)
125. Walters, R.W., Muhlrad, D., Garcia, J. and **R. Parker**. (2015) Differential effects of Ydj1 and Sis1 on Hsp70-mediated clearance of stress granules in *Saccharomyces cerevisiae*. **RNA** 21:1660-1671. [PMCID: PMC4536325](#)
126. Garcia, J.F. and **R. Parker**. (2015) MS2 coat proteins bound to yeast mRNAs block 5' to 3' degradation and trap mRNA decay products: implications for the localization of mRNAs by MS2-MCP system. **RNA** 8:1393-1395. [PMCID: PMC4509929](#)
127. Lin, Y., Protter, D.S.W., Rosen, M.K. and **R. Parker**. (2015) Formation and Maturation of Phase Separated Liquid Droplets by RNA Binding Proteins. **Molecular Cell** 60:208-219. [PMCID: PMC4609299](#)
128. Lasda, E. and **R. Parker**. (2016) Circular RNAs co-precipitate with extracellular vesicles: A possible mechanism for circRNA clearance. **PLOS ONE** 11(2):e0148407. [PMCID: PMC4743949](#)

Original Research Papers (continued)

129. Jain, S., Wheeler, J.R., Walters, R.W., Agrawal, A., Barsic, A. and **R. Parker**. (2016) ATPase-modulated stress granules contain a diverse proteome and substructure. **Cell** 164(3):487-498. [PMCID: PMC4733397](#)
130. Shukla, S., Schmidt, J.C., Goldfarb, K.C., Cech, T.R. and **R. Parker**. (2016) Inhibition of telomerase RNA decay rescues telomerase deficiency caused by dyskerin or PARN defects. **Nature Structural & Molecular Biology** 23:286-292. [PMCID: PMC4830462](#)
131. Garcia, J.F. and **R. Parker**. (2016) Ubiquitous accumulation of 3' mRNA decay fragments in *Saccharomyces cerevisiae* mRNAs with chromosomally integrated MS2 arrays. **RNA**, 22:657-659. [PMCID: PMC4836640](#)
132. Banani, S.F., Rice, A.M., Peebles, W.B., Lin, Y., Jain, S., **Parker, R.** and M.K. Rosen. (2016) Compositional control of phase-separated cellular bodies. **Cell**, 166:651-663. [PMCID: PMC4967043](#)
133. Eshleman, N., Liu, G., McGrath, K., **Parker, R.** and J.R. Buchan. (2016) Defects in THO/TREX-2 function cause accumulation of novel cytoplasmic mRNP granules that can be cleared by autophagy. **RNA** 22:1200-1214. [PMCID: PMC4931113](#)
134. Poornima, G., Shah, S., Vignesh, V., **Parker R.** and Rajyaguru, P.I. (2016) Arginine methylation promotes translation repression activity of eIF4G-binding protein, Scd6. **Nucleic Acids Research** 44:9358-9368. [PMCID: PMC5100564](#)
135. Wheeler, J.R., Matheny, T., Jain, S., Abrisch, R. and **R. Parker**. (2016) Distinct stages in stress granule assembly and disassembly. **eLife** Sep 7;5. pii: e18413 [PMCID: PMC5024549](#)
136. Harigaya, Y. and **R. Parker**. (2016) Codon optimality and mRNA decay. **Cell Research** 26:1269-1270. [PMCID: PMC5143416](#)
137. Harigaya, Y. and **R. Parker**. (2016) Analysis of the association between codon optimality and mRNA stability in *Schizosaccharomyces pombe*. **BMC Genomics** 17:895. [PMCID: PMC5101800](#)
138. Walters, R.W., Matheny, T., Mizoue, L.S., Rao, B.S., Muhlrad, D. and **R. Parker**. (2017) Identification of NAD⁺ capped mRNAs in *Saccharomyces cerevisiae*. **Proceedings of the National Academy of Science**. 114:480-485. [PMCID: PMC5255579](#)
139. Wheeler, J.R., Jain, S., Khong A. and **R. Parker**. (2017) Isolation of yeast and mammalian stress granule cores. **Methods** 126:12-17. [PMCID: PMC5924690](#)
140. Harigaya, Y. and **R. Parker**. (2017) The link between adjacent codon pairs and mRNA stability. **BMC Genomics** 18:364. [PMCID: PMC5424319](#)
141. Shukla, S. and **R. Parker**. (2017) PARN modulates Y RNA stability and its 3'-end formation. **Molecular and Cellular Biology** 37:e00264-17. [PMCID: PMC5615183](#)

Original Research Papers (continued)

142. Rao, B.S. and **R. Parker**. (2017) Numerous interactions act redundantly to assemble a tunable size of P bodies in *Saccharomyces cerevisiae*. **Proceedings of the National Academy of Science** 114(45):E9569-E9578. PMCID: PMC5692575
143. Khong, A., Matheny, T., Jain, S., Mitchell, S.F., Wheeler, J.R. and **R. Parker**. (2017) The stress granule transcriptome reveals principles of mRNA accumulation in stress granules. **Molecular Cell** 68:808-820. PMCID: PMC5728175
144. Tutucci, E., Vera, M., Biswas, J., Garcia, J., **Parker, R.** and R.H. Singer. (2018) An improved MS2 system for accurate reporting of the mRNA life cycle. **Nature Methods** 15(1):81-89. PMCID: PMC5843578
145. Protter, D.S.W., Rao, B.S., Van Treeck, B., Lin, Y., Mizoue, L., Rosen, M.K. and **R. Parker**. (2018) Intrinsically disordered regions can contribute promiscuous interactions to RNP granule assembly. **Cell Reports** 22(6):1401-1412. PMCID: PMC5824733
146. Khong, A., Jain, S., Matheny, T., Wheeler, J.R. and **R. Parker**. (2018) Isolation of mammalian stress granule cores for RNA-Seq analysis. **Methods** 137:49-54. PMCID: PMC5866748
147. Van Treeck, B., Protter, D.S.W., Matheny, T., Khong, A., Link, C.D. and **R. Parker**. (2018) RNA self-assembly contributes to stress granule formation and defining the stress granule transcriptome. **Proceedings of the National Academy of Science** 115(11):2734-2739. PMCID: PMC5856561
148. Bakthavachalu, B., Huelsmeier, J., Sudhakaran, I.P., Hillebrand, J., Singh, A., Petrauskas, A., Thiagarajan, D., Sankaranarayanan, M., Mizoue, L., Anderson, E.N., Pandey, U.B., Ross, E., VijayRaghavan, K., **Parker, R.** and M. Ramaswami. (2018) RNP-granule assembly via ataxin-2 disordered domains is required for long-term memory and neurodegeneration. **Neuron** 98(4):754-766. <https://doi.org/10.1016/j.neuron.2018.04.032>
149. Moon, S.L. and **R. Parker**. (2018) EIF2B2 mutations in vanishing white matter disease hypersuppress translation and delay recovery during the integrated stress response. **RNA** 24(6):841-852. PMCID: PMC5959252
150. Braselmann, E., Wierzba, A.J., Polaski, J.T., Chromiński, M., Holmes, Z.E., Hung, S.-T., Batan, D., Wheeler, J.R., **Parker, R.**, Jimenez, R., Gryko, D., Batey, R.T. and A.E. Palmer. (2018) A multicolor riboswitch-based platform for imaging of RNA in live mammalian cells. **Nature Chemical Biology** 14:964-971. PMCID: PMC6143402
151. Moon, S.L. and **R. Parker**. (2018) Analysis of eIF2B bodies and their relationships with stress granules and P-bodies. **Scientific Reports** 8(1):12264. PMCID: PMC6095920
152. Vogler, T.O., Wheeler, J.R., Nguyen, E.D., Hughes, M.P., Britson, K.A., Lester, E., Rao, B., Betta, N.D., Whitney, O.N., Ewachiw, T.E., Gomes, E., Shorter, J., Lloyd, T.E., Eisenberg, D.S., Taylor, J.P., Johnson, A.M., Olwin, B.B. and **R. Parker**. (2018) TDP-43 and RNA form amyloid-like myo-granules in regenerating muscle. **Nature** 563:508-513. PMCID: PMC6324568

Original Research Papers (continued)

153. Khong, A. and **R. Parker**. (2018) mRNP architecture in translating and stress conditions reveals an ordered pathway of mRNA compaction. **Journal of Cell Biology** Dec 3;217(12):4124-4140. [PMCID: PMC6279387](#)
154. Xing, W., Muhlrad, D. and **R. Parker**. (2018) A quantitative inventory of yeast P body proteins reveals principles of compositional specificity. **eLife** doi: 10.7554/eLife.56525
155. Moon, S.L., Morisaki, T., Khong, A., Lyon, K., **Parker. R.** and T.J. Stasevich. (2019) Multicolor single-molecule tracking of mRNA interactions with RNP granules. **Nature Cell Biology** February; 21(2):162–168. [PMCID: PMC6375083](#)
156. Shukla, S., Bjerke, G.A., Muhlrad, D., Yi, R. and **R. Parker**. (2019) The RNase PARN controls the levels of specific miRNAs that contribute to p53 regulation. **Molecular Cell** Mar 21;73(6):1204-1216. [PMCID: PMC6430647](#)
157. Fok, W.C., Shukla, S., Vessoni, A.T., Brenner, K.A., **Parker, R.**, Sturgeon, C.M. and L.F.Z. Batista. (2019) Posttranscriptional modulation of TERC by PAPD5 inhibition rescues hematopoietic development in dyskeratosis congenita. **Blood** 133(12):1308-1312. [PMCID: PMC6428664](#)
158. Tauber D. and **R. Parker**. (2019) 15-Deoxy- $\Delta^{12,14}$ -prostaglandin J2 promotes phosphorylation of eukaryotic initiation factor 2 α and activates the integrated stress response. **Journal of Biological Chemistry** Apr 19;294(16):6344-6352. [PMCID: PMC648127](#)
159. Baron, D.M., Matheny, T., Lin, Y.-C., Leszyk, J.D., Kenna, K., Gall, K.V., Santos, D.P., Tischbein, M., Funes, S., Hayward, L.J., Kiskinis, E., Landers, J.E., **Parker. R.**, Shaffer, S.A. and D.A. Bosco. (2019) Quantitative proteomics identifies proteins that resist translational repression and become dysregulated in ALS-FUS. **Human Molecular Genetics** 28(13):2143-2160. [PMCID PMC6586143](#)
160. Burke, J.M., Moon, S.L., Matheny, T. and **R. Parker**. (2019) RNase L reprograms translation by widespread mRNA turnover escaped by antiviral mRNAs. **Molecular Cell** 75:1203-1217. [PMCID: PMC6754297](#)
161. Decker, C.J., Steiner, H.R., Hoon-Hanks, L.L., Morrison, J.H., Haist, K.C., Stabell, A.C., Poeschla, E.M., Morrison, T.E., Stenglein, M.D., Sawyer, S.L. and **R. Parker**. (2019) dsRNA-Seq: Identification of viral infection by purifying and sequencing dsRNA. **Viruses** Oct 14;11(10):943. [PMCID: PMC6832592](#)
162. Matheny, T., Rao, B. and **R. Parker**. (2019) Transcriptome-wide comparison of stress granules and P-bodies reveals that translation plays a major role in RNA partitioning. **Molecular and Cellular Biology** 39(24): e00313-19 doi: 10.1128/MCB.00313-19 [PMCID: PMC6879202](#)
163. Tauber, D., Tauber, G., Khong, A., Van Treeck, B., Pelletier, J. and **R. Parker**. (2020) Modulation of RNA condensation by the DEAD-box protein eIF4A. **Cell** Feb 6;180:411-426.e16 [PMCID: PMC7194247](#)

Original Research Papers (continued)

164. Burke, J.M., Lester, E.T., Tauber, D. and **R. Parker**. (2020) RNase L promotes the formation of unique ribonucleoprotein granules distinct from stress granules. **Journal of Biological Chemistry** 295(6):1526-1438. [PMCID: PMC7008361](#)
165. Brocard, M., Iadevaia, V., Klein, P., Hall, B., Lewis, G., Lu, J., Burke, J., Wilcocks, M.M., **Parker, R.**, Goodfellow, I.G., Ruggieri, A. and N. Locker. (2020) Norovirus infection results in eIF2 α independent host translation shut-off and remodels the G3BP1 interactome evading stress granule formation. **PLOS Pathogens** Jan 6;16(1):E1008250 [PMCID: PMC6964919](#)
166. Cirillo, L., Cieren, A., Barbieri, S., Khong, A., Schwager, F., **Parker, R.** and M. Gotta. (2020) UBAP2L forms distinct cores that act in nucleating stress granules upstream of G3BP1. **Current Biology** 30(4):698-707.
167. Lee, J.E., Cathey, P.I. Wu, H., **Parker, R.** and G. Voeltz. (2020) Endoplasmic reticulum contact sites regulate the dynamics of membraneless organelles. **Science** 367(6477):eaay7108. doi: 10.1126/science.aay7108
168. Moon, S.L., Morisaki, T., Stasevich, T.J., and **R. Parker**. (2020) Coupling of translation quality control and mRNA targeting to stress granules. **Journal of Cell Biology** 219(8): e202004120 [PMCID: PMC7401812](#)
169. Shukla, S., Jeong, H.-C., **Parker, R.** and L.F.Z. Batista. (2020) Chemical inhibition of PAPD5/7 rescues telomerase function and hematopoiesis in dyskeratosis congenita. **Blood** 4(12):2717-2722. [PMCID: PMC7322949](#)
170. Xing, W., Muhlrad, D., **Parker, R.** and M.K. Rosen. (2020) A quantitative inventory of yeast P body proteins reveals principles of compositional and specificity. **eLife** 9:e56525 doi: [10.7554/eLife.56525](https://doi.org/10.7554/eLife.56525) [PMCID: PMC7373430](#)
171. Hoon-Hanks, L.L., Stöhr, A.C., Anderson, A.J., Evans, D.E., Nevarez, J.G., Raúl, D.E., Rodgers, C.P., Cross, S.T., Steiner, H.R., **Parker, R.** and M.D. Stenglein. (2020) Serpentivirus (nidovirus) and orthoreovirus coinfection in captive veiled chameleons (*Chamaeleo calyptratus*) with respiratory disease. **Viruses** 12(11):1329. [PMCID: PMC7699425](#)
172. Larremore, D.B., Wilder, B., Lester, E., Shehata, S., Burke, J.M., Hay, J.A., Tambe, M., Mina, M.J. and **R. Parker**. (2021) Test sensitivity is secondary to frequency and turnaround time for COVID-19 screening. **Science Advances** Jan; 7(1): eabd5393. [PMCID: PMC7775777](#)
173. Matheny, T., Van Treeck, B., Huynh, T.N. and **R. Parker**. (2021) RNA partitioning into stress granules is based on the summation of multiple interactions. **RNA** 27(2):174-189. doi: 10.1261/rna.078204.120. [PMCID: PMC7812873](#)

Original Research Papers (continued)

174. Yang, Q., Meyerson, N.R., Clark, S.K., Paige, C.L., Fattor, W.T., Gilchrist, A.R., Barbachano-Guerrero, A., Healy, B.G., Worden-Sapper, E.R., Wu, S.S., Muhlrad, D., Decker, C.J., Saldi, T.K., Lasda, E., Gonzales, P.K., Fink, M.R., Tat, K.L., Hager, C.R., Davis, J.C., Ozeroff, C.D., Brisson, G.R., McQueen, M.B., Leinwand, L., **Parker, R.**, and S. Sawyer. (2021) Saliva TwoStep for rapid detection of asymptomatic SARS-CoV-2 carriers. *eLife* 2021;10:e65113 doi: [10.7554/eLife.65113](https://doi.org/10.7554/eLife.65113) PMCID: PMC8057811
175. Lester, E., Ooi, F.K., Bakkar, N., Ayers, J., Woerman, A.L., Wheeler, J., Bowser, R., Carlson, G.A., Prusiner, S.B. and **R. Parker**. (2021) Tau aggregates are RNA-protein assemblies that mislocalize multiple nuclear speckle components. *Neuron* 109(10):1675-1691. doi.org/10.1016/j.neuron.2021.03.026 PMCID: PMC8141031
176. Burke, J.M., Gilchrist, A.R., Sawyer, S.L. and **R. Parker**. (2021) RNase L limits host and viral protein synthesis via inhibition of mRNA export. *Science Advances* Jun 4 7(23) eabh2479 DOI: 10.1126/sciadv.abh2479. PMCID: PMC8177694
177. Larremore, D., Toomre, D. and **R. Parker**. (2021) Modeling the effectiveness of olfactory testing to limit SARS-CoV-2 transmission. *Nature Communications* 12:3664. Jun 16;12(1):3664. doi: 10.1038/s41467-021-23315-5 PMCID: PMC8209051
178. Burke, J.M., St Clair, L.A., Perera, R. and **R. Parker**. (2021) SARS-CoV-2 infection triggers widespread host mRNA decay leading to an mRNA export block. *RNA* Nov 27(11):1318-1329 doi: [10.1261/rna.078923.121](https://doi.org/10.1261/rna.078923.121) PMCID: PMC8522697
179. Bjorkman, K., Saldi, T., Lasda, E., Bauer, L.C., Kovarik, J., Gonzales, P.K., Fink, M.R., Tat, K.L., Hager, C.R., Davis, J.C., Ozeroff, C.D., Brisson, G.R., Larremore, D.B., Leinwand, L.A., McQueen, M.B. and **R. Parker**. (2021) Higher viral load drives infrequent SARS-CoV-2 transmission between asymptomatic residence hall roommates. *Journal of Infectious Diseases* Oct 15 224(8):1316-1324 doi: [10.1093/infdis/jiab386](https://doi.org/10.1093/infdis/jiab386) PMCID: PMC8861368
180. Yang, Q., Saldi, T.K., Gonzales, P.K., Lasda, E., Decker, C.J., Tat, K.L., Fink, M.R., Hager, C.R., Davis, J.C., Ozeroff, C.D., Muhlrad, D., Clark, S.K., Fattor, W.T., Meyerson, N.R., Paige, C.L., Gilchrist, A.R., Barbachano-Guerrero, A., Worden-Sapper, E.R., Wu, S.S., Brisson, G.R., McQueen, M.B., Dowell, R.D., Leinwand, L., **Parker, R.** and S.L. Sawyer. (2021) Just 2% of SARS-CoV-2-positive individuals carry 90% of the virus circulating in communities. *Proceedings of the National Academy of Sciences* May 25, 118(21) E2104547118; doi.org/10.1073/pnas.2104547118 PMCID: PMC9166196
181. Corbet, G.A., Wheeler, J.R., **Parker, R.** and K. Weskamp. (2021) TDP43 ribonucleoprotein granules: physiologic function to pathologic aggregates. *RNA Biology* Oct 15;18(sup1):128-138. doi: [10.1080/15476286.2021.1963099](https://doi.org/10.1080/15476286.2021.1963099) PMCID: PMC8677035
182. Corbet, G.A., Burke, J.M. and **R. Parker**. (2021) ADAR1 limits stress granule formation through both translation-dependent and translation-independent mechanisms. *Journal of Cell Science* 134(17): ARTN jcs258783 doi: [10.1242/jcs.258783](https://doi.org/10.1242/jcs.258783) PMCID: PMC8445598

Original Research Papers (continued)

183. Reeves, K., Liebig, J., Feula, A., Saldi, T., Lasda, E., Johnson, W., Lilienfeld, J., Maggi, J., Pulley, K., Wilkerson, P.J., Real, B., Zak, G., Davis, J., Fink, M., Gonzales, P., Hager, C., Ozeroff, C., Tat, K., Alkire, M., Butler, C., Coe, E., Darby, J., Freeman, N., Heuer, H., Jones, J.R., Karr, M., Key, S., Maxwell, K., Nelson, L., Saldana, E., Shea, R., Salveson, L., Tomlinson, K., Vargas-Barriga, J., Vigil, B., Brisson, G., **Parker, R.**, Leinwand, L.A., Bjorkman, K. and C. Mansfeldt. (2021) High-resolution within-sewer SARS-CoV-2 surveillance facilitates informed intervention. **Water Research** Oct 1;204:117613. doi: 10.1016/j.watres.2021.117613. PMCID: PMC8402945
184. Decker, C.J., Burke, J.M., Mulvaney, P.K. and **R. Parker**. (2022) RNA is required for the maintenance of multiple cytoplasmic and nuclear membrane-less organelles. **EMBO Journal** May 2;41(9):e110137. doi: 10.15252/embj.2021110137 PMCID: PMC9058542
185. Khong, A., Matheny, T., Huynh, T.N., Babl, V. and **Roy Parker**. (2022) Limited effects of m⁶A modification on mRNA partitioning into stress granules. **Nature Communications** Jun 29;13(1):3735. doi:10.1038/s41467-022-31358-5 PMCID: PMC9243116
186. Bubar, K.M., Middleton, C.E., Bjorkman, K.K., **R. Parker** and D.B. Larremore. (2022) SARS-CoV-2 transmission and impacts of unvaccinated-only testing in populations of mixed vaccination status. **Nature Communications** May 19;13(1):2777 doi: 10.1038/s41467-022-30144-7 PMCID: PMC9120147
187. Iadevaia, V., Burke, J.M., Eke, L., Moller-Levet, C., **Parker, R.** and N. Locker. (2022) Novel stress granules-like structures are induced via a paracrine mechanism during viral infection. **Journal of Cell Science** Feb 15;135(4):jcs259194 doi: 10.1242/jcs.259194 PMCID: PMC8976915
188. Burke, J.M., Ripin, N., Ferretti, M.B., St. Clair, L.A., Worden-Sapper, E.R., Salgado, F., Sawyer, S.L., Perera, R., Lynch, K.W. and **R. Parker**. (2022) RNase L activation in the cytoplasm induces aberrant processing of mRNAs in the nucleus. **PLOS Pathogens** <https://doi.org/10.1371/journal.ppat.1010930> PMCID: PMC9651596
189. Wheeler, J.R., Whitney, O.N., Vogler, T.O., Nguyen, E.D., Pawlikowski, B., Lester, E., Cutler, A., Elston, T., Betta, N.D., Parker, K.R., Yost, K.E., Vogel, H., Rando, T.A., Chang, H.Y., Johnson, A.M., **Parker, R.**, and B.B. Olwin. (2022) RNA-binding proteins direct myogenic cell fate decisions. **eLife** Jun 13;11:e75844 doi: 10.7554/eLife.75844 PMCID: PMC9191894
190. Corbet, G.A., Burke, J.M., Bublitz, G.R., Tay, J.W. and **R. Parker**. (2022) dsRNA-induced condensation of antiviral proteins promotes PKR activation. **Proceedings of the National Academy of Sciences** Aug 16; 119(33) doi.org/10.1073/pnas.2204235119 PMCID: PMC9388085
191. Huynh, T.N., Shukla, S., Reigan, P., and **R. Parker**. (2022) Identification of PARN nuclease activity inhibitors by computational-based docking and high-throughput screening. **Scientific Reports** Mar 31. doi: [10.1038/s41598-023-32039-z](https://doi.org/10.1038/s41598-023-32039-z) PMCID: PMC10066322

Original Research Papers (continued)

192. Lester, E., Van Alstyne, M., McCann, K.L., Reddy, S., Cheng L., Kuo, J., Pratt, J., and **R. Parker**. (2023) Cytosolic condensates rich in polyserine define subcellular sites of tau aggregation. **Proceedings of the National Academy of Sciences** 120(3) doi.org/10.1073/pnas.2217759120
193. Jeong, H-C., Shukla, S., Fok, W.C., Huynh, T.H., Batista, L.F.Z., and **R. Parker**. (2023) USB1 is a miRNA deadenylase that regulates hematopoietic development. **Science** 379, 901-907. DOI: 10.1126/science.abj8379
194. Liang, Q., Chan, Y.-C., Toscano, J., Bjorkman, K.K., Leinwand, L.A., **R. Parker**, Nozik, E.S., Nesbitt, D.J., and J. Ye. (2023) Breath analysis by ultra-sensitive broadband laser spectroscopy detects SARS-CoV-2 infection. **Journal of Breath Research** Apr 5;17(3). doi: 10.1088/1752-7163/acc6e4
195. Currie, S.L., Xing, W., Muhlrad, D., Decker, C.J., **Parker, R.**, and M. K. Rosen. (2023) Quantitative reconstitution of yeast RNA processing bodies. **Proceedings of the National Academy of Sciences** <https://doi.org/10.1073/pnas.2214064120>
196. Shehata, S. and **R. Parker**. (2023) SARS-CoV-2 Nsp1 mediated mRNA degradation requires mRNA interaction with the ribosome. **RNA Biology** 20(1): 444–456. PMCID: PMC10332192
197. Yang, Q., Meyerson, N.R., Paige, C.L., Morrison, J.H., Clark, S.K., Fattor, W.T., Decker, C.J., Steiner, H.R., Lian, E., Larremore, D.B., Perera, R., Poeschla, E.M., **Parker, R.**, Dowell, R.D., and S. Sawyer. (2023) Human mRNA in Saliva can correctly identify individuals harboring acute infection. **mBio** <https://doi.org/10.1128/mbio.01712-23>
198. Ripin, N., Macedo de Vasconcelos, L., Ugay, D.A. and **R. Parker**. (2023) DDX6 modulates P-body and stress granule assembly composition and docking. (**submitted**)

BOOK CHAPTERS & INVITED REVIEWS

1. Heffron, F., Kostriken, R., Morita, C. and **R. Parker**. (1981) Tn3 encodes a site-specific recombination system: identification of essential sequences, genes, and the actual site of recombination. In: **Cold Spring Harbor Symposia on Quantitative Biology**, 45 Pt 1:259-268.
2. Jones, E., Zubenko, G., **Parker, R.**, Hemmings, B. and A. Hasik. (1981) Pleiotropic mutations of *Saccharomyces cerevisiae* which cause deficiency for proteinases and other vacuole enzymes. In: von Wettstein, D., Friis, J., Kielland-Brant, M., Stenderup, A. (eds) **Molecular Genetics in Yeast, Alfred Benzon Symposium**, Vol. 16, pp. 182-188.
3. Guthrie, C., Reidel, N., **Parker, R.**, Swerdlow, H. and B. Patterson. (1986) Genetic analyses of snRNAs and RNA processing in yeast. In: **Yeast Cell Biology, UCLA Symposia on Molecular Biology**, New Series, Vol. 33, pp. 301-321. New York: Alan R. Liss, Inc.

Book Chapters & Invited Reviews (continued)

4. Parker, R. and B. Patterson. (1987) Architecture of fungal introns: implications for spliceosome assembly. In: Inouye, M., and Dudock, B.S. (eds) **Molecular Biology of RNA: New Perspectives** pp. 133-149. New York: Academic Press.
5. Parker, R. (1989) Genetic methods for identification and characterization of RNA-RNA and RNA-protein interactions. **Methods in Enzymology** 180:510-517.
6. Jacobson, A., Herrick, D., Donahue, J., Parker, R. and S. Peltz. (1990) Regulation of mRNA stability in yeast. In: McCarthy, J.E.G., Puitte, M.F. (eds) **Post-Transcriptional Control of Gene Expression. NATO ASI Series**, H9:45-54.
7. Parker, R., Herrick, D., Peltz, S. and A. Jacobson. (1991) Measurement of mRNA decay rates in *Saccharomyces cerevisiae*. **Methods in Enzymology** 194:415-423.
8. Parker, R., Muhlrad, D., Deshler, J., Taylor, N. and J. Rossi. (1992) Ribozymes: Principles and designs for their use as antisense and therapeutic agents. In: Erickson, R.P. Izant, J.G. (eds) **Gene Regulation: Biology of Antisense RNA and DNA**. pp. 55-70. Raven Press, Ltd., New York.
9. Decker, C. and R. Parker. (1994) Mechanisms of mRNA degradation in eukaryotes. **Trends in Biochemical Sciences** 19:336-340.
10. Beelman, C. and R. Parker. (1995) Degradation of mRNA in eukaryotes. **Cell** 81:179-183.
11. Decker, C.J. and R. Parker. (1995) Diversity of cytoplasmic functions for the 3' untranslated region of eukaryotic transcripts. **Current Opinion in Cell Biology** 7:386-392.
12. Caponigro, G. and R. Parker. (1996) Mechanisms and Control of mRNA turnover in *Saccharomyces cerevisiae*. **Microbiological Reviews** 60:233-249. [PMCID: PMC239426](#)
13. Anderson, J.S.J. and R. Parker. (1996) RNA turnover: The helicase story unwinds. **Current Biology** 6:780-782.
14. LaGrandeur, T.E. and R. Parker. (1996) mRNA decapping activities and their biological roles. **Biochemie** 78:1049-1055.
15. Tharun, S. and R. Parker. (1997) Mechanisms of mRNA Turnover in Eukaryotic Cells. In: Morris, D.R. and Harford J.B. (eds) **mRNA Metabolism and Post-Transcriptional Gene Regulation**. pp. 181-199. New York, Wiley-Liss, Inc.
16. He, W. and R. Parker. (1999) Analysis of mRNA decay pathway in *Saccharomyces cerevisiae*. **Methods** 17:3-10.
17. Tharun, S. and R. Parker. (1999) Turnover of mRNA in Eukaryotic Cells. In: Barton, D., Nakanishi, K., Meth-Cohn, O. (eds) **Comprehensive Natural Products Chemistry, Volume 6: Prebiotic Chemistry, Molecular Fossils, Nucleosides, and RNA**. pp. 205-216. Amsterdam, New York, Elsevier Science, Ltd

Book Chapters & Invited Reviews (continued)

18. Hilleren, P. and **R. Parker**. (1999) mRNA surveillance in eukaryotes: Kinetic proofreading of proper translation termination as assessed by mRNP domain organization? **RNA** 5:711-719. [PMCID: PMC1369798](#)
19. van Hoof, A. and **R. Parker**. (1999) The exosome: A proteasome for RNA? **Cell** 99:347-350.
20. Hilleren, P. and **R. Parker**. (1999) Mechanisms of mRNA surveillance in eukaryotes. **Annual Review of Genetics** 3:229-260.
21. He, W. and **R. Parker**. (2000) Functions of Lsm proteins in mRNA degradation and splicing. **Current Opinion in Cell Biology** 12:346-350.
22. Tucker, M. and **R. Parker**. (2000) Mechanisms and control of mRNA decapping in *Saccharomyces cerevisiae*. **Annual Review of Biochemistry** 69:571-595.
23. Schwartz, D.C. and **R. Parker**. (2000) Interaction of mRNA Translation and mRNA Degradation in *Saccharomyces cerevisiae*. In: Sonenberg, N., Hershey, J.W.B., Matthews, M.B. (eds) **Translational Control of Gene Expression**. pp. 807-825. New York, Cold Spring Harbor Laboratory Press. DOI: 10.1101/087969618.39.807
24. Dunckley, T. and **R. Parker**. (2001) Yeast mRNA decapping enzyme. **Methods in Enzymology** 342:226-233.
25. Dunckley, T. and **R. Parker**. (2001) RNA Turnover. In: Brenner, S., Miller J.H. (eds) **Encyclopedia of Genetics**. Academic Press, NY, p1748-1751.
26. Wickens, M., Bernstein, D.S., Kimble, J. and **R. Parker**. (2002) A PUF family portrait: 3' UTR regulation as a way of life. **Trends in Genetics** 18:150-157.
27. van Hoof, A. and **R. Parker**. (2002) Messenger RNA degradation: Beginning at the end. **Current Biology** 12:R285-R287.
28. Steiger, M. and **R. Parker**. (2002) Analyzing mRNA decay in *Saccharomyces cerevisiae*. **Methods in Enzymology** 351:648-660.
29. Decker, C.J. and **R. Parker**. (2002) mRNA decay enzymes: Decappers conserved between yeast and mammals. **Proceeding of the National Academy of Science** 99:12512-12514. [PMCID: PMC130488](#)
30. **Parker, R.** and H. Song. (2004) The enzymes and control of eukaryotic mRNA turnover. **Nature Structural and Molecular Biology** 11:121-127.
31. Coller, J. and **R. Parker**. (2004) Eukaryotic mRNA decapping. **Annual Review of Biochemistry** 73:861-890.
32. Baker, K.E. and **R. Parker**. (2004) Nonsense-mediated mRNA decay: terminating erroneous gene expression. **Current Opinion Cell Biology** 16:293-299.

Book Chapters & Invited Reviews (continued)

33. Baker, K.E. and **R. Parker**. (2006) Features of nonsense-mediated mRNA decay in *Saccharomyces cerevisiae*. In: Maquat, L. (ed) **Nonsense-mediated mRNA Decay**. pp 3-13. Landes Biosciences, Texas.
34. Valencia-Sanchez, M., Liu, J., Hannon, G.J. and **R. Parker**. (2006) Control of translation and mRNA degradation by miRNAs and siRNAs. **Genes & Development** 20:515-524.
35. Decker, C.J. and **R. Parker**. (2006) CAR-1 and trailer hitch: Driving mRNP granule function at the ER? **Journal of Cell Biology** 173:159-163. PMCID: PMC2063806
36. Doma, M.K. and **R. Parker**. (2006) Revenge of the NRD: preferential degradation of nonfunctional eukaryotic rRNA. **Developmental Cell** 11:757-758.
37. **Parker, R.** and U. Sheth. (2007) P Bodies and the control of mRNA translation and degradation. **Molecular Cell** 25:635-646.
38. Doma, M.K. and **R. Parker**. (2007) RNA quality control in eukaryotes. **Cell** 131:660-668.
39. Buchan, J.R. and **R. Parker**. (2007) Molecular Biology: The two faces of miRNA. **Science** 318:1877-1878.
40. Beckham, C.J. and **R. Parker**. (2008) P Bodies, stress granules, and viral life cycles. **Cell Host & Microbe** 3:206-212. PMCID: PMC2396818
41. Hilliker, A. and **R. Parker**. (2008) "Stressed out? Make some modifications!" **Nature Cell Biology** 10:1129-1130. PMCID: PMC2803088
42. Nissan, T. and **R. Parker**. (2008) Analyzing P-bodies in *Saccharomyces cerevisiae*. **Methods in Enzymology** 448:507-520. PMCID: PMC2693489
43. Passos, D.O. and **R. Parker**. (2008) Analysis of cytoplasmic mRNA decay in *Saccharomyces cerevisiae*. **Methods in Enzymology** 448:409-427. PMCID: PMC2819139
44. Rajyaguru, P. and **R. Parker**. (2009) Cgh-1 and the control of maternal mRNAs. **Trends in Cell Biology** January 19, 19:24-28.
45. Balagopal, V. and **R. Parker**. (2009) Polysomes, P bodies and stress granules: states and fates of eukaryotic mRNAs. **Current Opinion in Cell Biology** 21:403-408. PMCID: PMC2740377
46. Swisher, K.D. and **R. Parker**. (2009) Related mechanisms for mRNA and rRNA quality control. **Molecular Cell** May 29, 34:401-402.
47. Thompson, D.M. and **R. Parker**. (2009) Stressing out over tRNA cleavage. **Cell** July 24, 138:215-219.
48. Buchan, J.R. and **R. Parker**. (2009) Eukaryotic stress granules: the ins and outs of translation. **Molecular Cell** 36:932-41. PMCID: PMC2813218

Book Chapters & Invited Reviews (continued)

49. Buchan, J.R., Nissan, T. and **R. Parker**. (2010) Analyzing P-Bodies and Stress Granules in *Saccharomyces cerevisiae*. **Methods in Enzymology** 470:619-640.
50. Yoon, J-H. and **R. Parker**. (2010) Coil-in-to snRNP assembly and Cajal bodies. **Nature Structural & Molecular Biology** 17:391-393. doi: 10.1038/nsmb0410-391
51. Luhtala, N. and **R. Parker**. (2010) T2 Family ribonucleases: ancient enzymes with diverse roles. **Trends in Biochemical Sciences** 35:253-259. PMCID: PMC2888479
52. Harigaya, Y. and **R. Parker**. (2010) No-go Decay: a quality control mechanism for RNA in translation. **WIREs RNA**, Wiley Interdisciplinary Reviews: RNA, 1:132-141. DOI:10.1002/wrna.17
53. **Parker, R.** (2012) RNA degradation in *Saccharomyces cerevisiae*. **Genetics** 191:671-702. doi: 10.1534/genetics.111.137265
54. Buchan, J.R., Capaldi, A.P. and **R. Parker**. (2012) TOR-tured yeast find a new way to stand the heat. **Molecular Cell** 47:155-157. doi: 10.1016/j.molcel.2012.07.005
55. Decker, C.J. and **R. Parker**. (2012) P-Bodies and stress granules: Possible roles in the control of translation and mRNA degradation. **Cold Spring Harbor Perspectives in Biology** Sep 1;4(9):a012286. doi: 10.1101/cshperspect.a012286 PMCID: PMC3428773
56. Rajyaguru, P. and **R. Parker**. (2012) RGG motif proteins: Modulators of mRNA functional states. **Cell Cycle** 11:2594-2599. PMCID: PMC3873214
57. Jain S., **Parker R.** (2013) The Discovery and Analysis of P Bodies. In: Chan E., Fritzler M. (eds) **Ten Years of Progress in GW/P Body Research. Advances in Experimental Medicine and Biology**, vol 768. Springer, New York, NY
58. Ramaswami, M., Taylor J.P., and **R. Parker**. (2013) Altered Ribostasis: RNA-protein granules in degenerative disorders. **Cell** 154:727-736. PMCID: PMC3811119
59. Gilbert, J.A., Ball, M., Blainey, P., Blaser, M.J., Bohannan, B.J.M., Bunge, J., Dominguez-Bello, M.G., Epstein, S., Fierer, N., Gevers, D., Grikscheit, T., Hamdan, L., Harvey, J., Huttenhower, C., Kirkup, B., Kong, H.H., Lauber, C., Lemon, K., Lych, S., Martin, L., Mello, C., Palma, J., **Parker, R.**, Petrosino, J., Segre, J.A., Voshall, L., Yi, R. and R. Knight. (2014) Meeting Report for the 1st Skin Microbiota Workshop, Boulder, CO October 15-16 2012. **Standards in Genomic Sciences** 9:13:1-7. PMCID: PMC4334105
60. Walters, R. and **R. Parker**. (2014) Quality control: Is there quality control of localized mRNAs? **Journal of Cell Biology** 204:863-868. DOI: 10.1083/jcb.201401059
61. Mitchell, S.F. and **R. Parker**. (2014) Principles and properties of 3ukaryotic mRNPs. **Molecular Cell** 54:547-558.
62. Harigaya, Y. and **R. Parker**. (2014) Fragile X mental retardation protein and the ribosome. **Molecular Cell** 3:330-332.

Book Chapters & Invited Reviews (continued)

63. Lasda, E.L. and **R. Parker**. (2014) Circular RNAs: Diversity of form and function. **RNA** 20:1829-1842. [PMCID: PMC4238349](#)
64. Schwartz, J.C., Cech, T.R. and **R. Parker**. (2015) Biochemical properties and biological functions of FET Proteins. **Annual Review of Biochemistry** 84:355-379. doi: 10.1146/annurev-biochem-060614-034325. Epub 2014 Dec 8.
65. Mitchell S.F., **Parker R.** (2015) In Vivo cross-linking followed by PolyA Enrichment to Identify Yeast mRNA Binding Proteins. In: Boudvillain M. (eds) **RNA Remodeling Proteins. Methods in Molecular Biology**, vol 1259. Humana Press, New York, NY.
66. Walters, R.W. and **R. Parker**. (2015) Coupling of ribostasis and proteostasis: Hsp70 proteins in mRNA metabolism. **Trends in Biochemical Sciences** 40:552-559. [PMCID: PMC4584423](#)
67. Mitchell, S.F. and **R. Parker**. (2015) Modifications on translation initiation. **Cell** 163:796-798.
68. Mitchell S.F., **Parker R.** (2016) Identification of Endogenous mRNA-Binding Proteins in Yeast Using Crosslinking and PolyA Enrichment. In: Lin R.-J. (eds) **RNA-Protein Complexes and Interactions. Methods in Molecular Biology**, vol 1421. Humana Press, New York, NY
69. Prottier, D.S.W. and **R. Parker**. (2016) Principles and properties of stress granules. **Trends in Cell Biology** 26:668-679. [PMCID: PMC4993645](#)
70. Shukla, S. and **R. Parker** (2016) Hypo- and hyper-assembly diseases of RNA-protein complexes. **Trends in Molecular Medicine** 22:615-628. [PMCID: PMC4925306](#)
71. Moon, S.L., Sonenberg, N. and **R. Parker**. (2018) Neuronal regulation of eIF2 α function in health and neurological disorders. **Trends in Molecular Medicine** 24(6):575-589.
72. Van Treeck, B. and **R. Parker**. (2018) Emerging Roles for Intermolecular RNA-RNA Interactions in RNP Assemblies. **Cell** 174(4):791-802.
73. Mittag, T. and **R. Parker**. (2018) Multiple modes of protein-protein interactions promote RNP granule assembly. **Journal of Molecular Biology** 430(23):4636-4649. [PMCID: PMC6204294](#)
74. Lester, E. and **R. Parker**. (2018) The Tau of Nuclear-Cytoplasmic Transport. **Neuron** 99(5):869-871.
75. Van Treeck, B. and **R. Parker**. (2019) Principles of stress granules revealed by imaging approaches. **Cold Spring Harbor Perspectives in Biology** Feb 1;11(2). [PMCID: PMC63690856](#)
76. Cutler, A.A., Ewachiw, T.E., Corbet, G.A., **Parker, R.** and B.B. Olwin. (2019) Myo-granules connect physiology and pathophysiology. **Journal of Experimental Neuroscience** Apr 12;13:1179069519842157 eCollection 2019. [PMCID: PMC6463236](#)

Book Chapters & Invited Reviews (continued)

77. Corbet G.A. and **R. Parker**. (2019) RNP Granule Formation: Lessons from P-Bodies and Stress Granules. *Cold Spring Harb Symp Quant Biol*. 2019;84:203-215. doi: 10.1101/sqb.2019.84.040329. Epub 2020 Jun 1.
78. Khong, A. and **R. Parker**. (2020) The Landscape of Eukaryotic mRNPs. *RNA* Mar;26(3):229-239. PMCID: PMC7025503
79. Tauber, D., Tauber, G. and **R. Parker**. (2020) Mechanisms and Regulation of RNA Condensation. *Trends in Biochemical Sciences* May;45(9):764-778. PMCID: PMC7211619
80. Mina, M.J., **Parker, R.** and D. Larremore. (2020) Rethinking Covid-19 Test Sensitivity — A Strategy for Containment. *New England Journal of Medicine* Nov 26;383(22):e120. doi: 10.1056/NEJMp2025631. Epub 2020 Sep 30.
81. Weskamp, K., Olwin, B. and **R. Parker**. (2021) Post-transcriptional regulation in skeletal muscle development, repair, and disease. *Trends in Molecular Medicine* May;27(5):469-481.
82. Pratt, J., Lester, E. and **R. Parker**. (2021) Could SARS-CoV-2 cause tauopathy? *The Lancet Neurology*, Jul;20(7):506. [https://doi.org/10.1016/S1474-4422\(21\)00168-X](https://doi.org/10.1016/S1474-4422(21)00168-X). PMCID: PMC8292136
83. Batista, L.F.Z., Dokal, I., and **R. Parker**. (2022) Telomere biology disorders: time for moving towards the clinic? *Trends in Molecular Medicine* October;28(10):882-891. <https://doi.org/10.1016/j.molmed.2022.08.001>
84. Ripin, N. and **R. Parker**. (2022) Are stress granules the RNA analogs of misfolded protein aggregates? *RNA* 28(1)67-75. doi: [10.1261/rna.079000.121](https://doi.org/10.1261/rna.079000.121) PMCID: PMC8675284
85. Corbet, G.A., Burke, J.M. and **R. Parker**. (2022) Nucleic acid-protein condensates in innate immune signaling. *EMBO Journal* 42:e111870 <https://doi.org/10.15252/embj.2022111870>
86. Tollervey, D. and **R. Parker**. (2022) Christine Guthrie: Splicing genetics and mentorship into the RNA world. *Proceedings of the National Academy of Sciences* November, 119(47) e2216884119, <https://www.pnas.org/doi/10.1073/pnas.2216884119>
87. Lester, E. and **R. Parker**. (2023) Tau, RNA, and RNA binding proteins: complex interactions in health and neurodegenerative diseases. *The Neuroscientist* Mar 9;10738584231154551. doi: 10.1177/10738584231154551
88. Huynh, T.N. and **R. Parker**. (2023) The PARN, TOE1, and USB1 RNA deadenylases and their roles in non-coding RNA regulation. *Journal of Biological Chemistry* Aug 4;105139. doi: [10.1016/j.jbc.2023.105139](https://doi.org/10.1016/j.jbc.2023.105139)
89. Ripin, N. and **R. Parker**. (2023) Formation, Function, and Pathology of RNP Granules. *Cell* 186(22): 4737-4756. <https://doi.org/10.1016/j.cell.2023.09.006>